

Five New Cassane-Type Diterpenes from *Caesalpinia crista*

by Zheng-Yi Yang^{a)1)}, Yin-Hua Yin^{a)1)}, and Li-Hong Hu^{*a)1)}

^{a)} Shanghai Research Center for Modernization of Traditional Chinese Medicine, Shanghai Institute of Materia Medica, Chinese Academy of Sciences, 199 Guoshoujing Road, Shanghai 201203, P. R. China

^{b)} School of Pharmacy, East China University of Science and Technology, 130 Meilong Road, Shanghai 200237, P. R. China (phone/fax: +86-21-50272221; e-mail: simmhulh@mail.shcnc.ac.cn)

Five new cassane-type diterpenes, caesalpinista A (**1**), caesalpinista B (**2**), caesaljapin B (**3**), caesaljapin C (**4**), and caesalpinilinn (**5**) were isolated from the MeOH extract of the seeds of *Caesalpinia crista*. Their structures were elucidated by the analysis of their 1D- and 2D-NMR spectra.

Introduction. – *Caesalpinia crista* L. (Fabaceae) is a well-known medicinal plant widely distributed in tropical and subtropical regions of Southeast Asia. This plant is locally known as ‘Ka-Lain’ in Myanmar, and its seeds are used as an anthelmintic, antipyretic, anti-inflammatory, and antimalarial agent [1]. In Indonesia, it is known as ‘Bagore’, and a decoction of its roots has been used as a tonic and for the treatment of rheumatism and backache [2]. As a member of the genus *Caesalpinia*, it is a rich source of cassane-type furanoditerpenes and is reported to have antimalarial [3][4], antiviral [5], and anticancer activities [6]. We have chemically investigated the seeds of *Caesalpinia crista* which resulted in the isolation of five new cassane diterpenes (**1–5**; see Fig. 1). In this paper, we report the structure elucidation of these cassane-type diterpenes.

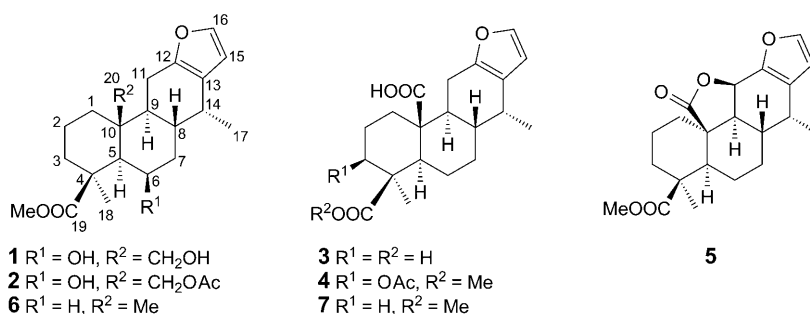


Fig. 1. The structures of compounds **1–7**

Results and Discussion. – Compound **1** was isolated as a colorless amorphous solid and its molecular formula was determined to be C₂₁H₃₀O₅ by HR-EI-MS. The IR

¹⁾ The authors contributed equally to this work.

absorptions at 3423 and 1718 cm^{-1} indicated the presence of OH and CO groups, respectively. The $^1\text{H-NMR}$ spectrum (Table 1) of **1** displayed signals corresponding to two H-atoms of a 1,2-disubstituted furan ring ($\delta(\text{H})$ 7.24, 6.22), a sharp *singlet* due to a MeOCO group ($\delta(\text{H})$ 3.71), one tertiary Me group ($\delta(\text{H})$ 1.57), and one Me *doublet* signal at $\delta(\text{H})$ 0.90 (*d*, $J = 6.9$). The $^{13}\text{C-NMR}$ spectrum (Table 2) of **1** showed four olefinic C-atoms ($\delta(\text{C})$ 149.6, 140.6, 122.4, and 109.9) and two O-substituted C-atoms ($\delta(\text{C})$ 69.1, 64.6), together with one ester CO C-atom ($\delta(\text{C})$ 179.6). The $^1\text{H-}$ and $^{13}\text{C-NMR}$ data (Tables 1 and 2) were similar to those of deoxycsaesaldekarin C (**6**) [7], except that a *multiplet* signal of $\text{CH}_2(6)$ ($\delta(\text{H})$ 1.56) was replaced by a CH–O group at $\delta(\text{H})$ 3.89 and one tertiary Me group at $\delta(\text{H})$ 0.93 (Me(20)) was replaced by an CH_2O group at $\delta(\text{H})$ 4.25 and 3.60 (each *d*, $J = 12.6$). In the HMBC and HSQC spectra, we observed the correlations of the CH–O group ($\delta(\text{H})$ 3.89) with C(5) ($\delta(\text{C})$ 51.3) and C(7) ($\delta(\text{C})$ 39.6), as well as of the CH_2O group ($\delta(\text{H})$ 4.25, 3.62) with C(10) ($\delta(\text{C})$ 41.2), which indicated the CH–O group was located at C(6) and the CH_2O group was at C(20).

Table 1. $^1\text{H-NMR}$ Data (500 MHz) of **1–3** in CDCl_3 or CD_3OD . $\delta(\text{H})$ in ppm, J in Hz^{a} .

	1 ^b	2 ^b	3 ^c
$\text{CH}_2(1)$	1.94 (<i>d</i> , $J = 12.9$), 1.08–1.20 (<i>m</i>)	2.42 (<i>d</i> , $J = 12.7$), 0.98–1.02 (<i>m</i>)	2.17–2.21 (<i>m</i>), 1.00–1.04 (<i>m</i>)
$\text{CH}_2(2)$	1.48–1.55 (<i>m</i>), 1.32–1.45 (<i>m</i>)	1.60–1.64 (<i>m</i>), 1.40–1.46 (<i>m</i>)	1.88–1.92 (<i>m</i>), 1.60–1.64 (<i>m</i>)
$\text{CH}_2(3)$	1.73–1.88 (<i>m</i>), 1.58–1.62 (<i>m</i>)	1.76–1.82 (<i>m</i>), 1.64–1.68 (<i>m</i>)	1.80–1.84 (<i>m</i>), 1.57–1.61 (<i>m</i>)
H–C(5)	2.02 (<i>br. s</i>)	1.98 (<i>br. s</i>)	1.98 (<i>br. s</i>)
H–C(6) or $\text{CH}_2(6)$	3.89 (<i>br. s</i>)	3.98 (<i>br. s</i>)	2.16–2.20 (<i>m</i>), 1.20–1.24 (<i>m</i>)
$\text{CH}_2(7)$	1.62–1.66 (<i>m</i>)	1.68–1.72 (<i>m</i>)	2.74–2.78 (<i>m</i>), 1.40–1.44 (<i>m</i>)
H–C(8)	2.42–2.52 (<i>m</i>)	2.18–2.24 (<i>m</i>)	2.46 (<i>d</i> , $J = 13.4$)
H–C(9)	1.68–1.72 (<i>m</i>)	1.72–1.76 (<i>m</i>)	1.58–1.62 (<i>m</i>)
$\text{CH}_2(11)$	2.72–2.76 (<i>m</i>), 2.52–2.60 (<i>m</i>)	2.75 (<i>dd</i> , $J = 16.4, 6.1$), 2.50 (<i>dd</i> , $J = 16.4, 11.1$)	2.68 (<i>dd</i> , $J = 16.4, 6.1$), 2.10 (<i>dd</i> , $J = 16.4, 11.1$)
H–C(14)	2.68–2.72 (<i>m</i>)	2.58–2.64 (<i>m</i>)	2.57–2.61 (<i>m</i>)
H–C(15)	6.22 (<i>s</i>)	6.20 (<i>s</i>)	6.18 (<i>s</i>)
H–C(16)	7.24 (<i>s</i>)	7.25 (<i>s</i>)	7.22 (<i>s</i>)
Me(17)	0.90 (<i>d</i> , $J = 6.9$)	0.98 (<i>d</i> , $J = 6.9$)	0.98 (<i>d</i> , $J = 7.4$)
Me(18)	1.57 (<i>s</i>)	1.58 (<i>s</i>)	1.08 (<i>s</i>)
$\text{CH}_2(20)$	4.25 (<i>d</i> , $J = 12.6$), 3.60 (<i>d</i> , $J = 12.6$)	4.98 (<i>d</i> , $J = 13.5$), 4.30 (<i>d</i> , $J = 13.5$)	
COOMe	3.71 (<i>s</i>)	3.70 (<i>s</i>)	
OAc		2.04 (<i>s</i>)	

^a) Assignments were made using HSQC and HMBC data. ^b) In CDCl_3 . ^c) In CD_3OD .

The relative configuration of **1** was determined on the basis of ROESY correlations. The ROESY correlations of Me(18) ($\delta(\text{H})$ 1.57) with H–C(5) ($\delta(\text{H})$ 2.02) and H–C(6) ($\delta(\text{H})$ 3.89), of $\text{CH}_2(20)$ ($\delta(\text{H})$ 4.25, 3.60) with $\text{H}_{\text{ax}}\text{-C}(2)$ ($\delta(\text{H})$ 1.32–1.45)

Table 2. ^{13}C -NMR Data (125 MHz) of **1–5** in CDCl_3 or CD_3OD . $\delta(\text{C})$ in ppm.

	1 ^{a)}	2 ^{a)}	3 ^{b)}	4 ^{a)}	5 ^{a)}
C(1)	38.6	34.8	37.6	33.5	33.3
C(2)	18.9	18.6	21.1	22.8	18.0
C(3)	39.6	38.2	38.6	77.0	37.0
C(4)	48.6	48.2	49.2	52.1	47.2
C(5)	51.3	51.1	51.7	50.2	45.6
C(6)	69.1	69.6	25.0	24.5	26.5
C(7)	39.6	40.0	31.9	29.8	29.5
C(8)	32.8	31.2	37.7	35.2	38.4
C(9)	45.5	45.4	45.3	43.4	47.0
C(10)	41.2	41.1	50.1	47.8	50.2
C(11)	22.5	22.9	25.6	24.3	68.2
C(12)	149.6	149.3	150.1	147.8	144.7
C(13)	122.4	121.9	124.3	122.7	130.4
C(14)	31.4	31.0	33.2	31.1	31.7
C(15)	109.9	109.5	111.0	109.5	109.2
C(16)	140.6	140.4	142.3	140.7	143.8
C(17)	17.2	17.8	18.0	17.1	15.1
C(18)	18.8	18.5	16.4	10.5	16.9
C(19)	179.6	179.2	182.4	175.7	178.9
C(20)	64.6	64.6	179.1	180.1	175.9
COOMe	52.4	52.2		52.3	52.0
OCOMe		170.9		170.2	
OCOMe		21.2		21.0	

^{a)} Recorded in CDCl_3 at 125 MHz. ^{b)} Recorded in CD_3OD at 125 MHz.

and $\text{H}_{\text{ax}}\text{-C}(11)$ ($\delta(\text{H})$ 2.52–2.60) indicated that rings *A* and *B* have a chair conformation with a *trans*-fused ring junction. On the other hand, ROESY correlations of $\text{H-C}(6)$ ($\delta(\text{H})$ 3.89) with $\text{H-C}(5)$ ($\delta(\text{H})$ 2.02) and $\text{Me}(18)$ ($\delta(\text{H})$ 1.57) indicated that the OH substituent at C(6) was β -oriented and that the Me(18) was α -oriented (Fig. 2). From these spectral evidences, the structure of **1** was determined and named as caesalpinista A.

Compound **2** showed the molecular ion peak at m/z 404.2199 ($\text{C}_{23}\text{H}_{32}\text{O}_6^+$; calc. 404.2190) in the HR-EI-MS. The ^1H - and ^{13}C -NMR spectral data (Tables 1 and 2) revealed that **2** had the same cassane-type carbon skeleton as **1**. The only difference was the presence of an additional AcO group ($\delta(\text{H})$ 2.04 and $\delta(\text{C})$ 170.9, 21.2). The $\text{CH}_2(20)\text{-O}$ group of **2** was shifted downfield from $\delta(\text{H})$ 4.25 and 3.60 (each *d*, $J = 12.6$) to $\delta(\text{H})$ 4.98 and 4.30 (each *d*, $J = 13.5$) as a result of the acetylation of the OH group. In addition, a significant HMBC between $\text{CH}_2(20)$ and the AcO CO group ($\delta(\text{C})$ 170.9) further confirmed the location of the AcO group at C(20). Thus, the structure of **2** was determined and named as caesalpinista B.

Compound **3** was deduced as $\text{C}_{20}\text{H}_{26}\text{O}_5$ by HR-EI-MS analysis (M^+ , m/z 346.1780; calc. 346.1784). The ^1H - and ^{13}C -NMR spectral data (Tables 1 and 2) of **3** were closely related to those of caesaljapin (**7**) [8]. The only difference between them was the lack of a MeOCO group at C(19). Therefore, the structure of **3** was determined and named as caesaljapin B.

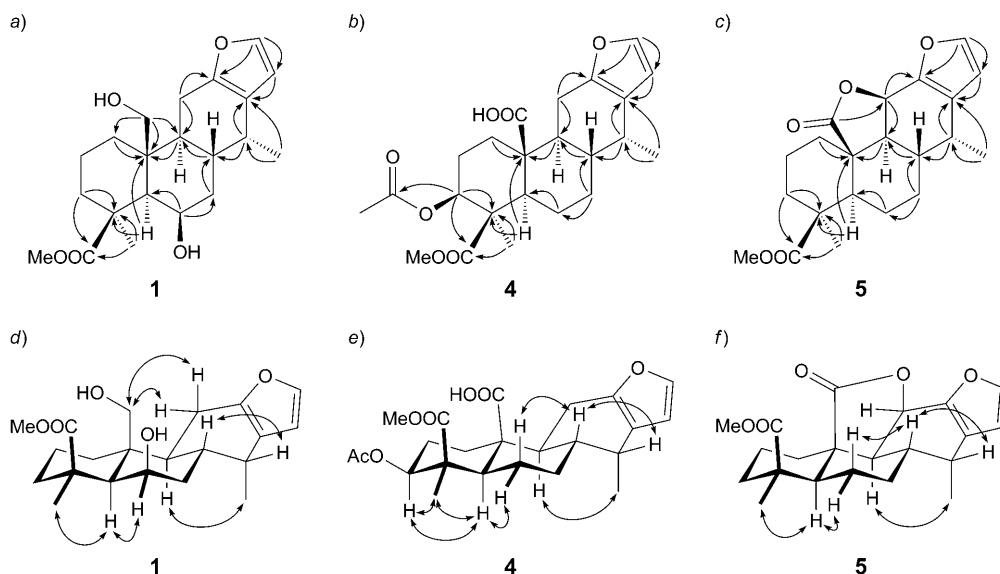


Fig. 2. Major HMBC data of compounds a) **1**, b) **4**, and c) **5**; important ROESY cross-peaks of compounds d) **1**, e) **4**, and f) **5**

Compound **4** was isolated as a colorless amorphous solid and its molecular formula was determined to be $C_{23}H_{30}O_7$ by HR-EI-MS. The 1H - and ^{13}C -NMR spectral data (Tables 3 and 2) also revealed the same cassane-type skeleton as caesaljapin (**7**). The 1H -NMR spectral data exhibited a CH–O group at $\delta(H)$ 5.21 (*dd*, $J = 9.7, 7.0$) and an

Table 3. 1H -NMR Data (500 MHz) of **4**–**5** in $CDCl_3$. δ (H) in ppm, J in Hz^a)

	4	5
CH ₂ (1)	2.55 (<i>d</i> , $J = 13.6$), 1.20–1.25 (<i>m</i>)	2.50 (<i>d</i> , $J = 13.1$), 1.44–1.50 (<i>m</i>)
CH ₂ (2)	2.28–2.36 (<i>m</i>), 1.00–1.10 (<i>m</i>)	1.64–1.68 (<i>m</i>), 1.52–1.58 (<i>m</i>)
H–C(3) or CH ₂ (3)	5.21 (<i>dd</i> , $J = 9.7, 7.0$)	1.68–1.72 (<i>m</i>)
H–C(5)	1.94 (<i>br. s</i>)	2.16 (<i>dd</i> , $J = 7.6, 4.7$)
CH ₂ (6)	1.88–1.92 (<i>m</i>)	1.36–1.42 (<i>m</i>)
CH ₂ (7)	1.70–1.78 (<i>m</i>), 1.35–1.45 (<i>m</i>)	1.58–1.64 (<i>m</i>), 1.42–1.46 (<i>m</i>)
H–C(8)	2.18–2.22 (<i>m</i>)	1.78–1.88 (<i>m</i>)
H–C(9)	1.60–1.68 (<i>m</i>)	2.06 (<i>dd</i> , $J = 12.0, 3.6$)
CH ₂ (11) or H–C(11)	2.80 (<i>dd</i> , $J = 16.3, 5.9$), 2.10–2.15 (<i>m</i>)	5.38 (<i>d</i> , $J = 3.6$)
H–C(14)	2.60–2.68 (<i>m</i>)	2.62–2.68 (<i>m</i>)
H–C(15)	6.15 (<i>s</i>)	6.25 (<i>s</i>)
H–C(16)	7.18 (<i>s</i>)	7.38 (<i>s</i>)
Me(17)	0.98 (<i>d</i> , $J = 6.9$)	0.98 (<i>d</i> , $J = 7.2$)
Me(18)	1.13 (<i>s</i>)	1.52 (<i>s</i>)
COOMe	3.68 (<i>s</i>)	
OAc	1.98 (<i>s</i>)	

^a) Assignments were made using HSQC and HMBC data.

AcO group ($\delta(\text{H})$ 1.98). The CH–O group showed HMBC correlations with C(4) ($\delta(\text{C})$ 52.1), C(18) ($\delta(\text{C})$ 10.5), C(19) ($\delta(\text{C})$ 175.7), the AcO group ($\delta(\text{C})$ 170.2), and C(2) ($\delta(\text{C})$ 22.8), confirming an AcO group at C(3). The ROESY correlations of Me(18) ($\delta(\text{H})$ 1.13) with H–C(5) ($\delta(\text{H})$ 1.94) and H–C(3) ($\delta(\text{H})$ 5.20) indicated the AcO group at C(3) to be in β -axial orientation (Fig. 2). Thus, the structure of **4** was determined and named as caesaljapin C.

Compound **5** showed the molecular ion peak at m/z 358.1886 ($\text{C}_{21}\text{H}_{26}\text{O}_5^+$; calc. 358.1881) in HR-EI-MS. The ^1H - and ^{13}C -NMR spectral data (see Tables 3 and 2) also revealed that **5** had the similar cassane-type skeleton as caesaljapin. The ^1H -NMR spectral data exhibited a CH–O group at $\delta(\text{H})$ 5.38 ($d, J = 3.6$). The CH–O group showed HMBC correlations with C(9) ($\delta(\text{C})$ 47.0), C(12) ($\delta(\text{C})$ 144.7), C(13) ($\delta(\text{C})$ 130.4), and C(20) ($\delta(\text{C})$ 175.9), confirming that the location of the CH–O group was at C(11) and it was linked with C(20) by an ester bond. The configuration at C(11) was determined as β -OH by the cross-peak between H–C(9) ($\delta(\text{H})$ 2.06, $dd, J = 12.0, 3.6$) and H–C(11) ($\delta(\text{H})$ 5.38, $d, J = 3.6$) in ROESY experiments and the small coupling constant between them (Fig. 2). Thus, compound **5** was determined and named as caesalpinilinn.

Accordingly, as a result of this investigation, the structures of five new compounds from *Caesalpinia crista* were identified.

Experimental Part

General. All solvents used were of chemical grade (Shanghai Chemical Plant). TLC: precoated silica-gel GF₂₅₄ plates (Qingdao Haiyang Chemical Plant). Column chromatography (CC): silica gel (SiO_2 ; 200–300 mesh); MCI Gel CHP20P (75–150 μm ; Mitsubishi Kasei Chemical Industries); C₁₈ reverse-phased SiO_2 (20–45 μm , Fuji Silysia Chemical Ltd.); Sephadex LH-20 (Pharmacia). Optical rotations: Perkin-Elmer model 341 polarimeter. IR Spectra: Bio-Rad-FT-IR spectrophotometer, ν in cm^{-1} . NMR spectra: Bruker AMX-500 spectrometer (500 MHz for ^1H and 125 MHz for ^{13}C); conventional pulse sequences for ROESY, HSQC and HMBC; 200 ms mixing time for ROESY; chemical shifts δ in ppm, J in Hz; CDCl_3 and CD_3OD solns. HR-EI-MS: positive mode; Bruker Atex III spectrometer.

Plant Material. The seeds of *Caesalpinia crista* were collected in Zhejiang Province, P. R. China in May, 2006. A voucher specimen of the plant was identified by Mr. Jin-Gui Shen and deposited at the Shanghai Research Center for Modernization of Traditional Chinese Medicine, Shanghai Institute of Materia Medica, Shanghai.

Extraction and Isolation. The dried and powered seeds of *Caesalpinia crista* (10.0 kg) were extracted successively with MeOH at r.t. (3×5 l) overnight. The conc. extract was partitioned between CHCl_3 and H_2O . Evaporation of CHCl_3 left a dark residue (150 g). The residue was subjected to MCI gel CHP 20P CC, eluted with MeOH/ H_2O (30:70, 70:60, 90:50, 100:0) to yield four subfractions (Fr. A–D). Fr. B (35 g) was subjected to SiO_2 (200–300 mesh), eluted with hexane/acetone (10:1, 5:1, 3:1, 1:1, acetone) to yield three subfractions (Fr. B-1–B-3). Fr. B-1 (3.6 g) was chromatographed by RP-18 flash CC, eluted with MeOH/ H_2O (40:60) to afford compounds **1** (18 mg) and **2** (15 mg). Fr. B-2 (0.8 g) was passed through a Sephadex LH-20 column, eluted with MeOH to give **5** (13 mg). Fr. B-3 (7.1 g) was subjected to MCI gel CHP 20P CC, eluted with MeOH/ H_2O (60:40 and 70:30) to afford compound **3** (98 mg) and **4** (12 mg).

Caesalpinista A (= Methyl (4S,4aR,5R,6aS,7R,11aS,11bS)-1,2,3,4,4a,5,6,6a,7,11,11a,11b-Dodecahydro-5-hydroxy-11b-(hydroxymethyl)-4,7-dimethylphenanthro[3,2-b]furan-4-carboxylate; **1**). White amorphous powder. $[\alpha]_{\text{D}}^{20} = +0.076$ ($c = 0.105$, MeOH). IR (KBr): 3423, 2929, 1718, 1637, 1072. ^1H - and ^{13}C -NMR (CDCl_3): Tables 1 and 2. HR-ESI-MS: 362.2093 (M^+ , $\text{C}_{21}\text{H}_{30}\text{O}_5^+$; calc. 362.2089).

Caesalpinista B (= Methyl (4*S*,4*aR*,5*R*,6*aS*,7*R*,11*aS*,11*bS*)-11*b*-[(Acetoxy)methyl]-1,2,3,4,4*a*,5,6,6*a*,7,11,11*a*,11*b*-dodecahydro-5-hydroxy-4,7-dimethylphenanthro[3,2-*b*]furan-4-carboxylate; **2**). White amorphous powder. $[\alpha]_{\text{D}}^{20} = +0.114$ ($c = 0.12$, MeOH). IR (KBr): 3416, 2921, 1746, 1638, 1062. ^1H - and ^{13}C -NMR (CDCl_3): Tables 1 and 2. HR-ESI-MS: 404.2199 (M^+ , $\text{C}_{23}\text{H}_{32}\text{O}_5^+$; calc. 404.2190).

Caesaljin B (= (4*S*,4*aS*,6*aS*,7*R*,11*aS*,11*bS*)-2,3,4,4*a*,5,6,6*a*,7,11,11*a*-Decahydro-4,7-dimethylphenanthro[3,2-*b*]furan-4,11*b*(1*H*)-dicarboxylic Acid; **3**). White amorphous powder. $[\alpha]_{\text{D}}^{20} = +0.104$ ($c = 0.12$, MeOH). IR (KBr): 3411, 2968, 2927, 2862, 2642, 2239, 2077, 1693, 1646, 1454, 1276, 1242, 1053, 738. ^1H - and ^{13}C -NMR (CD_3OD): Tables 1 and 2. HR-ESI-MS: 346.1780 (M^+ , $\text{C}_{20}\text{H}_{26}\text{O}_5^+$; calc. 346.1784).

Caesaljin C (= (3*S*,4*R*,4*aR*,6*aS*,7*R*,11*aS*,11*bS*)-3-(Acetyloxy)-2,3,4,4*a*,5,6,6*a*,7,11,11*a*-dodecahydro-4-(methoxycarbonyl)-4,7-dimethylphenanthro[3,2-*b*]furan-11*b*(1*H*)-carboxylic Acid; **4**). White amorphous powder. $[\alpha]_{\text{D}}^{20} = +0.084$ ($c = 0.105$, MeOH). IR (KBr): 3428, 2956, 2931, 2869, 1735, 1646, 1448, 1369, 1249, 1143, 1028. ^1H - and ^{13}C -NMR (CDCl_3): Tables 3 and 2. HR-ESI-MS: 418.1992 (M^+ , $\text{C}_{23}\text{H}_{30}\text{O}_7^+$; calc. 418.1985).

Caesalpinilinn (= Methyl (4*S*,4*aS*,6*aS*,7*R*,10*bR*,12*aR*,12*bS*)-2,3,4,4*a*,5,6,6*a*,7,10*b*,12*b*-Decahydro-4,7-dimethyl-12-oxo-1*H*-phenanthro[5,4*a*-*bc*:6,7-*b'*]difuran-4-carboxylate; **5**). White amorphous powder. $[\alpha]_{\text{D}}^{20} = +0.003$ ($c = 0.1$, MeOH). IR (KBr): 3434, 2935, 2863, 1762, 1724, 1452, 1738, 1253, 1124, 1072, 937. ^1H - and ^{13}C -NMR (CDCl_3): Tables 3 and 2. HR-ESI-MS: 358.1886 (M^+ , $\text{C}_{21}\text{H}_{26}\text{O}_5^+$; calc. 358.1881).

Financial support by the National Natural Science Foundation of China (NNSF; No. 30371679, 30623008), Chinese National High-tech R&D Programs (2006AA020602, 2006AA02Z156 and 2007AA02Z100) and Shanghai Science and Technology Commission (06DZ22028) is gratefully acknowledged.

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Received June 17, 2008